SYNTHESIS OF POTENTIAL ECDYSTEROID PRECURSORS FROM Δ^{7+22} STEROLS

Udo Hedtmann,* Kurt Hobert,* Tsenka Milkova,b and Peter Welzel* *

*Fakultät für Chemie der Ruhr-Universität Postfach 102148, D-4630 Bochum (FRG)

^bBulgarian Academy of Sciences, Institute of Organic Chemistry, 1113 Sofia (Bulgaria)

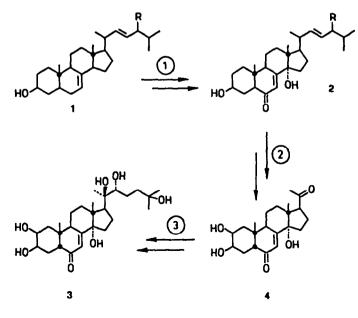
(Received in Germany 29 December 1987)

<u>Abstract</u> - Two routes for the conversion of 5α -cholest-7-ene-32-ol (11) into 7, which has the typical ecdysteroid substitution pattern in rings B, C, and D, have been developed. Making use of one of the methods, 5,6-dihydroergosterol (31) was converted into the 38, possessing all the functionalities required for the synthesis of naturally occurring ecdysteroids.

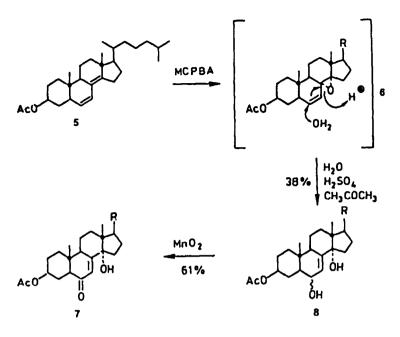
Introduction

Whereas Δ^3 -sterols such as cholesterol, β -sitosterol, and campesterol play an important rôle as starting materials for the synthesis of biologically active steroids,^{1,2} little efforts have been devoted to developing methods for the use of compounds of type 1 (R=H, CH₃, C_{2H₅}) as steroidal raw materials. Δ^7 -sterois (examples of which are chondrillasterol³ or spinasterol³) have been isolated from many sources (higher⁴ and lower plants,⁵ marine organisms⁴). Since the last three years we embarked on a project aimed at the synthesis of ecdysteroids such as 20-hydroxyecdysone (3) using the general sequence $1 \rightarrow 2 \rightarrow 4 \rightarrow 3$ (Scheme 1). We have already published a new method for the conversion of poststeron (4) into 3.⁷ Here we wish to outline the successful execution of part 1 of our synthetic plan (Scheme 1, $1 \rightarrow 2$).

We based our investigations on work of Wada,^{β} who has found that 5 α -cholesta-6,8(14)-diene (5) on reaction with m-chloroperbenzoic acid gives the unsaturated epoxide 6. Under acidic conditions 6 hydrolyses to provide the enedici 8, which



Scheme 1



Scheme 2

on oxidation with manganese dioxide provides 7 with the typical ecdysteroid substitution pattern in rings B, C, and D (Scheme 2).

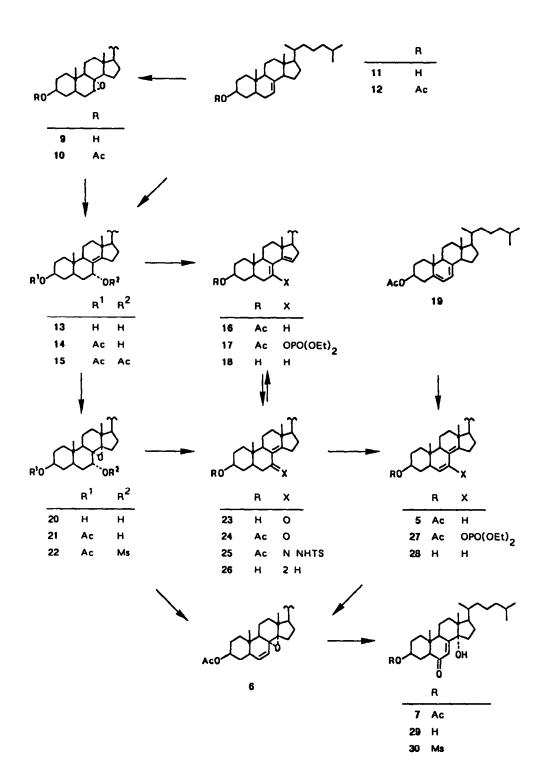
Conversion of 11 and 19 into 7.

The overall yield reported by Wada⁸ for the formation of 7 starting from 5, was 23%. We found that the yield was considerably increased when after peracid oxidation the intermediate 6 was immediately oxidized with sodium dichromate in acetic acid. 7 was then obtained in an overall yield of 68%.

Dienes of type 5 are easily available by rearrangement of 5,7-dienes such as 19.^{9,10} (Scheme 3). Combined with the oxidation methodology discussed above this constitutes an efficient access of ecdysteroid-like substituted compounds such as 7 using 5,7-dienes as starting materials.¹¹

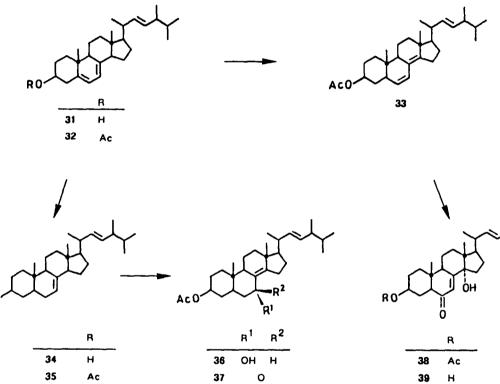
For Δ '-sterols such as 11 the situation is more complicated. Allyl alcohol 14 which is prepared from 12 via selenium dioxide oxidation¹² or by acid-catalyzed rearrangement of epoxide 10 under carefully controlled conditions (Fieser performed the reaction in chloroform solution¹³, we obtained best results with 0.01 molar p-toluenesulfonic acid in dimethoxyethane (DME): 76%) was shown to exclusively the unwanted diene 16 on elimination with furnish thionyl chloride.12,14 On attempted conversion of 14 into the tosylate and the mesylate, respectively, we also isolated only 16.15 The same result was obtained when acetate 15 was submitted to Trost's eliminination conditions¹⁶ (heating of 15 with molybdenum hexacarbonyl to 110°C in toluene solution).

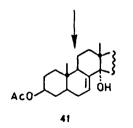
In order to bypass these difficulties, **11** was converted to **24** by a) exidation with two equivalents of m-chloroperbenzoic acid to give $20, 1^{7,18}$ b) acid-catalyzed rearrangement to give $23, 1^{7,18}$ and c) acetylation. On reaction with tosylhydrazine **24** furnished two isomeric hydrazones **25** (rather sensitive compounds), the configurations of which were not determined.¹⁹ In their reactivity towards methyl lithium (Shapiro reaction ²⁰) the two tosylhydrazones differed somewhat. One isomer reacted faster (24h at 20°C) and yielded diene **28** (the desired precursor of **7**, vide supra) and its isomer **18** in a **4**.5:1 ratio (combined yield: 76%). For the Shapiro reaction of the other hydrazone an elevated temperature had to be chosen

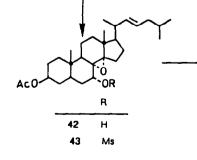


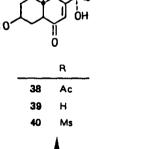
(20h at 40°C). In this case, 28 and 18 were formed in a 1.6:1 ratio. It should be pointed out that diene 28 could not be obtained from 24 by using of the Ireland method.²¹ Treatment of **24** with a) lithium diisopropylamide and b) diethyl chlorophosphate furnished dienol phosphate 17 rather than 27. Reduction of 17 with lithium in ethylamine led to the formation of 26 (75%).

In a different set of experiments 12 was oxidized with m-chloroperbenzoic acid (2 equivalents) according to Fieser and Djerassi^{17,18} providing **21**. Mesylation followed by elimination (by heating of 22 in diazabicyclononane) gave 6 which was directly oxidized with sodium dichromate in acetic acid whereby the final product 7 was obtained in 85% yield.









44

Ac O.

Scheme 4

In conclusion: We have developed two routes from a 1^7 -sterol to 7. The first sequence $(11 \rightarrow 20 \rightarrow 23 \rightarrow 24 \rightarrow 25 \rightarrow 28 \rightarrow 5 \rightarrow 7)$ comprises eight steps with an overall yield of 18^{8} .²² In a second, much more practical five-step route $(11 \rightarrow 12 \rightarrow 21 \rightarrow 6 \rightarrow 7)$ an overall yield of 40% was achieved.

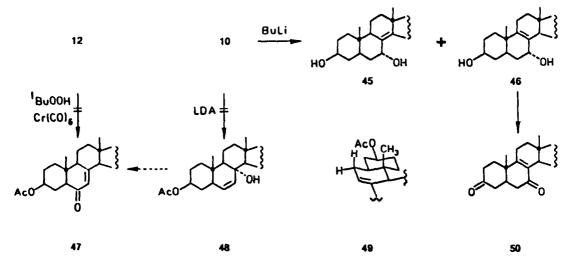
Conversion of 31 and 34 into 39

As a model for 7,22-dienic sterols, dihydroergosterol^{23,24} was chosen. The Fieser-Djerassi oxidation^{17,18} of 35 to give 42 (49% yield) had to be performed under carefully selected conditions otherwise the side-chain double bond reacted as well.²⁵ 42 was also obtained from 35 by a) selenium dioxide oxidation²⁶ to furnish a 3:1 mixture of 36 and a compound which we presume to be 41, since it slowly rearranged into 36, and b) oxidation of 36 with pyridinium dichromate (PDC), which led to 42 along with 37. 42 was converted to 44 by a) mesylation and b) elimination, essentially as described above. Finally, oxidation with sodium dichromate in acetic acid provided 38. 38 was also prepared from 32 by a) double bond isomerization, b) m-chloroperbenzoic acid oxidation, and c) oxidation with sodium dichromate in acetic acid, as described for the conversion of 19 into 7.

From 38 and 7, the corresponding mesylates 40 and 30, respectively, have been prepared by acetate hydrolysis and mesylation. 30 and 40 have a functionality in ring A which is well-suited for the introduction of the ecdysteroid 2,3-diol system by elimination and hydroxylation.²⁷

Addendum

At the outset of our work we intended to make use of Pearson's allylic oxidation method.²⁸ However, all attempts to prepare 47 by oxidation of 12 with tbutylhydroperoxide/chromium hexacarbonyl failed.²⁹ We believe that 68-H, which has to be abstracted in the oxidation process³⁰, is for sterical reasons not accessible (see Scheme 5, formula 49). We also tried to rearrange epoxide 10 into allyl alcohol 48³¹ which we hoped to oxidize to 45 with pyridinium chlorochromate.³² Again, all attempts met with no success. We were unable to rearrange 10 with lithium disopropylamide, and the reaction of 10 with butyllithium furnished allyl alcohols 45 and 46 which are of little use in the present context.



Scheme 5

EXPERIMENTAL

<u>General</u>

All O₂- or moisture-sensitive reactions were performed in oven-dried glassware under a positive pressure of argon. Sensitive liquids and solutions were transferred by syringe and were introduced into the reaction flask through rubber septa. Usual work-up means partitioning the reaction mixture between water and an organic solvent (given in paranthesis), drying the combined organic solutions over Na2SO4 and removal of the solvent by distillation in vacuo at 40°C, using a rotatory evaporator. The instrumentation used was: ¹H NMR: WP-80 (Bruker); AM-400 (Bruker); ¹³C NMR: AM-400 (Bruker); IR: Perkin Elmer 257; MS: MAT-731 and MAT-CH-5 (Finnigan); HPLC: High-pressure chromatography using pump system model 6000 (Waters Associates Inc.), UV-detector LC-3 (Pye Unicam), stainless steel columns 25.0 cm x 0.4 cm (analytical) or 25.0 cm x 2 cm (preparative), stationary phase and eluent are given in parenthesis; MPLC: Medium-pressure chromatography using 31.0 cm x 2.5 cm glass tubes, silica gel Grace (50 μ m), Duramat pump (CFG); Thomachrom UV detector (Reichelt).

7α.8-Epoxy-5α.8α-cholestan-3β-yl acetate (10).

10 was prepared from **12** ²⁴ using the procedure of Fieser et al. ¹³: yield 75%.-M.p. 95-96°C (from methanol, lit., ¹³ 96-97°C).- IR (CCl₄): 1740 cm⁻¹(CO).- ¹H NMR (80 MHz, CDCl₃): δ = 0.74 (s, CH₃-18), 0.84 (s, CH₃-19), 2.02 (s, OAc), 3.30 (m, W_{1/2}=5 Hz, 7-H), 4.45-4.85 (m, 3-H) .- C₂₉H₄₈O₃ (444.6), MS: m/z (%) = 444 (28, M⁺), 426 (24), 411 (8), 366 (9), 351 (11) 313 (29), 120 (100)

<u>5a-Cholest-8(14)-ene-38,7a-diol 3-acetate (14).33</u>

To a solution of **10** (1.0 g, 2.25 mmol) in DME (25 ml) a 0.01 m solution of ptoluenesulfonic acid in diethylether (0.6 ml) was added. The mixture was stirred at 25°C for 2 h. The acid was then neutralized by addition of solid K₂CO₃ Filtration, solvent evaporation and LC (hexanes-ethyl acetate-NEt₃ 7:1:0.01) gave **14** (720 mg, 72%) and **16** (180 mg, 18%).- M.p. 113-115°C (from methanol).- IR (CCl₄): 3600, 3450 (OH), 1725 cm⁻¹ (CO).- ¹H NMR (80 MHz, CDCl₃): δ = 0.71 (s, CH₃-18), 0.87 (s, CH₃-19), 2.02 (s, OAc), 4.53 (m, W_{1/2}=6 Hz, 7-H), 4.50-4.90 (m, 3-H).- C_{29H46}O₃ (444.6), MS: m/z (%) = 444 (100, M⁺), 426 (24), 351 (13), 331 (13), 313 (42), 290 (45).

Reaction of 15 with molybdenum hexacarbonyl.

To a solution of **15** (18 mg, 0.04 mmol, prepared from **5** as described by Fieser ¹²) O,N-bistrimethylsilylacetamide (0.01 ml) was added. The mixture was stirred at 110°C (sealed reaction flask) for 15 min. After cooling to 25°C, molybdenum hexacarbonyl (2 mg, 0.008 mmol) was added. The mixture was stirred at 110°C for 16 h. Filtration through silica gel (0.5 g, hexanes-ethyl acetate 10:1) and solvent evaporation gave **16**.- M.p. 84-86°C (from methanol, lit., ³⁴ 85-86°C). When the reaction was performed at 80°C, **15** was recovered.

8,14-Epoxy-5α,8α-cholestan-3β,7α-diol (20).

To a solution of **10** (2.0 g, 5.2 mmol) in CH_2Cl_2 (50 ml) at 25°C MCPBA (2.5 g, 15.0 mmol) was added in small portions. The mixture was kept at 4-6°C for 160 h. After solvent evaporation the residue was redissolved in diethylether (50 ml). Usual work-up and MPLC (hexanes-ethyl acetate 3:2) furnished **20** (1.4 g, 66%) along with small amounts of the corresponding 8,9-epoxide (0.3 g, 16%).- M.p. 186-187°C (from ethanol, lit.,¹² 186-187°C).- IR (CHCl₃): 3600-3300 cm⁻¹ (OH).- ¹H NMR (80 MHz, CDCl₃): $\delta = 0.80-1.00$ (CH₃ signals), 2.20-2.40 (m, OH), 3.35-3.80 (m, 3-H), 3.52 (W_{1/2}=6 Hz, 7-H).- C_{27H46}O₃ (418.6), MS: m/z (%) = 418 (40, M⁺), 403 (26), 400 (6), 385 (7), 369 (9), 305 (37), 287 (60), 269 (29), 107 (100).

<u>3B-Acetoxy-5 α -cholest-8(14)-en-7-one (24).</u>

A solution of 20 (1.9 g, 4.5 mmol) in ethanol (70 ml) and conc. HCl (3.5 ml) was refluxed for 2 h. Water (100 ml) was added and the mixture was cooled to 0°C. A crystalline fraction was obtained by filtration and the filtrate was extracted with diethylether and the ethereal solution was combined with the crystalline product (dissolved in 100 ml of ether), washed with saturated aq. NaHCO3 and water and dried over Na₂SO₄. After solvent evaporation the residue was acetylated (pyridine (20 ml), acetic anhydride (6 ml), 16 h at 20°C). Evaporation to dryness (after addition of toluene) and MPLC (hexanes-ethyl acetate 3:2) furnished 24 (1.5 g, 75%) along with small amounts of the corresponding 8(14)-en-15-one isomer.¹⁷ M.p. 140-142°C (from ethanol, lit.,¹⁷ 141-142°C).- IR (CHCl₃): 1720, 1665 cm⁻¹.-¹H NMR (80 MHz, CDCl₃): δ = 0.81 (s, CH₃-18), 0.91 (s, CH₃-19), 2.01 (s, OAC), 4.45-4.95 (3-H).- C_{29H46O3} (442.4), MS: m/z (%) = 442 (M⁺, 100), 357 (65), 329 (29), 276 (59).

<u>38-Acetoxy-5a-cholesta-7.14-dien-7-vl diethvl phosphate (17).</u>

To a LDA solution ³⁵ (0.33 mmol in THF (1 ml)) at -78°C a solution of **24** (80 mg, 0.18 mmol) in dry THF (3.3 ml) was added. After being allowed to warm to ambient temperature the mixture was stirred for 1.5 h. Diethyl chlorophosphate (173 mg, 1.0 mmol) was added and the mixture was stirred for 1 h at 20°C. Usual work-up (diethylether) and MPLC (hexanes-ethyl acetate 3:1) gave **17** (45 mg, 43%) and recovered **24** (15mg, 19%).- IR (CCl₄): 1725 (CO), 1625 cm⁻¹ (C=C).- ¹H NMR (400 MHz, CDCl₃): δ = 0.69 (s, CH₃-18), 0.84 (d, J=6 Hz, CH₃-26, CH₃-27), 0.86 (s, CH₃-19), 0.92 (d, J=6 Hz, CH₃-21), 1.32 (m, CH₃-CH₂-O), 2.01 (s, OAc), 2.16 (m, 1 H), 2.52-2.62 and 2.75-2.86 (2 m' s, CH₂-6), 4.12 (m, CH₃-CH₂-O), 4.67-4.77 (m, 3-H), 5.18 (m, 15-H).- C_{33H35}O₆P (578.7), MS: m/z (%) = 578 (85, M⁺), 563 (5), 518 (8), 465 (10), 440 (9), 424 (70), 380 (25), 364 (100), 349 (39), 339 (27), 311 (57), 251 (81).

Reduction of 17.

17 was prepared from 24 (100 mg. 0.23 mmol) as described above. To the crude reaction mixture at -78°C tert. butanol (0,07 ml) was added. This mixture was introduced dropwise at -60°C into a solution of lithium (14 mg, 2,0 mmol) in ethylamine (2 ml). After being stirred for 1 h at -60°C the mixture was allowed to warm to ambient temperature and was quenched with 1:1 THF/water. Evaporation of ethylamine, usual work-up (diethylether) and LC (hexanes-ethyl acetate 5:1) furnished 5a-cholest-7-en-38-ol (26)(30 mg, 35%).- M.p. 117-119°C (from ethanol, lit., ³⁶ 119-119.5°C).

<u>38-Acetoxy-5g-cholest-8(14)-en-7-one-(p-toluene-sulfonylhydrazone) (25), two</u> stereoisomers.

A mixture of 24 (400 mg, 0.9 mmol), p-toluene-sulfonylhydrazine (310 mg, 1.6 mmol), CH_2Cl_2 (10 ml), and conc. HCl (3 drops) was stirred at 40°C (sealed reaction flask) for 5 h. After being cooled to ambient temperature to the mixture was added solid K_2CO_3 . Filtration, solvent evaporation, and MPLC (hexanes-ethyl acetate 6:1) gave the unpolar hydrazone (300 mg, 55 %) and then the polar isomer (230 mg, 42%).

Unpolar isomer: M.p. 129-132°C (from diisobutylether).- IR (CHCl₃): 3300, 3200 (NH), 1720 (CO), 1600 cm⁻¹ (C=C).- ¹H NMR (80 MHz, CDCl₃): $\delta = 0.78$ (s, CH₃-18), 0.90 (s, CH₃-19), 2.02 (s, OAc), 2.43 (s, Ar-CH₃), 4.35-4.85 (m, 3-H), 7.30 and 7.77 (Ar-H).- CD (ethanol): max (ϵ) = 275 (+4.10), 269 (+3.56), 249 (-3.33), 232 (+2.50), 209 (-15.77), 197 nm (+35.10).- C₃ ϵ Hs₄N₂O₄S (610.8), MS: m/z (ϵ) = 610 (0.4, M⁺), 454 (45), 453.3455 (100, Calc for C₂₉H₄₅N₂O₂: 453.3481), 452 (75), 437 (8), 426 (8), 314 (25).

Polar isomer: IR (CHCl₃): 3300, 3200 (NH), 1720 (CO), 1600 cm⁻¹ (C=C).- ¹H NMR (80 MHz, CDCl₃): δ = 0.72 (s, CH₃-18), 0.84 (s, CH₃-19), 2.03 (s, OAc), 2.43

(s, Ar-CH₃), 4.40-4.90 (m, 3-H), 7.30 and 7.85 (Ar-H).- CD (ethanol): max (ϵ) = 275 (+11.62) 270 (+11.70), 218 (-8.02), 198 nm (+7.37).- C₃₆H₅₄N₂O₄S (610.8) MS: m/z (\Re) = 454 (47), 453.3446 (100, Calc for C₂₉H₄₅N₂O₂: 453.3481), 452 (72), 437 (8), 426 (12), 314 (25).

Shapiro reaction of 25.

a) unpolar isomer: To a solution of **25** (unpolar isomer, 50 mg, 0.082 mmol) in dry diethylether (2 ml) a 1.6 M solution of methyllithium in diethylether (0.26 ml, 0.41 mmol) was added and the mixture was stirred for 24 h at 20°C. After addition of water (2 ml) and usual work-up (diethylether) LC (hexanes-ethyl acetate 5:1) gave a mixture of **18** and **28** ³⁷ 24 mg, 76%; ratio 1:4.5 (from the intensities of the olefinic proton signals (80 MHz)).

b) polar isomer: Using the above procedure from **25** (polar isomer) a 1.6:1 mixture of **28** and **18** was obtained in 72 % yield.

8,14-Epoxy-5a,8a-cholestan-3ß,7a-diol 3-acetate 7-mesylate (22).

To a solution of 21 (350 mg, 0.76 mmol) in dry pyridine (6 ml) methanesulfonyl chloride (0.43 ml, 0.43 mmol) was added. The mixture was stirred at 40°C for 16 h. Solvent evaporation (by codistillation with toluene) and LC (hexanes-ethyl 87%).- M.p. (dec.) (from acetate 6:1) gave **22** (335 mg, 111-112°C hexanes/ethylacetate).- IR (CCl4): 1735 (CO), 1360, 1170 cm⁻¹ (SO₂).- ¹H NMR (80 MHz, CDCl₃): $\delta = 0.80-1.00$ (CH₃ signals), 2.02 (s, OAc), 3.10 (s, SO₂CH₃), 4.50-4.90 (m, 3-H, 7-H).- MS: m/z (%) = 442 (15), 424 (26), 409 (50), 349 (19), 327 (12), 311 (40), 301 (29), 43 (100).- (Found: C, 66.80; H, 9.33; CloH5006S (538.8) requires C, 66.87; H, 9.35).

8.14-Epoxy-5a.8a-cholest-6-en-3B-yl acetate (6).

A solution of 22 (514 mg, 0.96 mmol) in DBN (7.5 ml) was stirred at 80°C for 38 h. Usual work-up (diethylether) and MPLC (hexanes-ethyl acetate-NEt₃ 9:1:0.05) gave 6 (312 mg, 75%).- M.p. 91-92°C/115-120°C (from hexanes-CH₂Cl₂).- IR (CCl₄): 1735 (CO), 1620 cm⁻¹ (C=C).- ¹H NMR (80 MHz, CDCl₃): $\delta = 0.80-1.00$ (CH₃ signals), 2.02 (s, OAc), 4.50-4.90 (m, 3-H), 5.22 (6-H), and 5.60 (7-H, AB-part of an ABX-system, J7,6=9 Hz, J₆,5=3 Hz, J₇,s=2 Hz).- MS: m/z (%) = 442 (4, M*), 424 (4), 382 (11), 364 (5), 349 (9), 329 (100).- (Found: C, 78.70; H, 10.50; C_{2.9}H_{4.6}O₃ (442.6) requires C, 78.68; H, 10.47).

<u>3B-Acetoxy-14-hydroxy-5g-cholest-7-en-6-one (7).</u>

a) From 5: A solution of 5 (207 mg, 0.49 mmol) and MCPBA (110 mg, 0.55 mmol) in diethylether (15 ml) was kept at 4-6°C for 24 h. Then a solution of sodium dichromate (160mg, 0.61 mmol) in 96 % acetic acid (4 ml) was added and the mixture was stirred at 20°C for 15 min. Usual work-up (diethylether) and LC (hexanes-ethyl acetate 3:1) gave 20 (152 mg, 68 %).- M.p. 185-186°C (from methanol, lit.,⁶ 186-187°C).

b) From 6 : To a solution of 6 (50 mg, 0.11 mmol) in diethylether (5 ml) at $20 \circ C$ a solution of sodium dichromate (40 mg, 0.15 mmol) in 96 % acetic acid (1 ml) was added. The mixture was stirred at $20 \circ C$ for 30 min and then diluted with diethylether (15 ml). Usual work-up and LC (hexanes-ethyl acetate 5:2) gave 7 (43 mg, 83 %).

14-Hydroxy-38-mesyloxy-5g-cholest-7-en-6-one (30).

To a solution of **29** (49 mg, 0.12 mmol, prepared from **7** as described in ref. ¹¹) in dry pyridine (2 ml), methanesulfonyl chloride (0.02 ml, 0.2 mmol) was added and the mixture was stirred at 25°C for 3 h. Usual work-up (diethylether) and LC (hexanes-ethyl acetate 5:2) gave **30** (42 mg, 69 %).- M.p. 157-159°C (from hexanesethyl acetate).- IR (CHCl₃): 3590 (OH), 1670 (unsat. CO), 1365, 1170 cm⁻¹ (SO₂).-¹H NMR (80 MHz, CDCl₃): $\delta = 0.79$ (s, CH₃-18), 0.89 (s, CH₃-19), 3.02 (s, SO₂CH₃), 4.40-4.85 (m, 3-H), 5.90 (d, J=2 Hz, 7-H).~ MS: m/z (%) = 494.3064 (2, M⁺, Calc for C₂₈H₄₆O₅S : 494.3066), 476 (25), 461 (32), 398 (18), 380 (38), 365 (89), 363 (100), 349 (20), 326 (20), 267 (64).

Selenium dioxide oxidation of 35.

To a solution of 35 (150 mg, 0.33 mmol, prepared from ergosterol 31 by hydrogenation using Wilkinson's catalyst to give 34 and subsequent acetylation ³⁸) in dioxane (6 ml) a solution of SeO_2 (39 mg, 0.33 mmol) in 1:9 water-dioxane (6 ml) was added. The mixture was stirred under reflux for 10 min and after cooling it was poured into ice water. Usual work-up (diethylether) gave a crude product which was redissolved in diethylether (25 ml) and filtered. Solvent evaporation and LC (hexanes-diethylether, gradient 3:1 - 0:100) gave 36 (85 mg, 55%) and 41 (16 mg, 10%).

5a-Ergosta-8(14).22-diene-38.7a-diol 3-acetate (36).

M.p. 129-131°C (from pentane, lit., 3^9 125-126°C).- IR (CCl₄): 3600, 3550-3350 (OH), 1730 cm⁻¹ (CO).- ¹H NMR (80 MHz, CDCl₃): 8 = 0.65-1.00 (CH₃ signals), 2.00 (s, OAc), 4.47 (m, W_{1/2}=6 Hz, 7-H), 4.50-4.85 (m, 3-H), 5.12-5.25 (m, 22-H, 23-H).- MS: m/z (%) = 456.3597 (100, M*, Calc for C₃oH₄sO₃: 456.3603), 438 (9), 423 (4), 395 (2), 331 (32), 313 (14), 312 (15), 302 (8), 253 (16).

5g-Brgosta-7.22-diene-38.14g-diol 3-acetate (41).

M.p. 198-200°C (from diethylether/pentane).⁴°- IR (CC14): 3600, 3550-3400 (OH), 1730 cm⁻¹ (CO).- ¹H NMR (80 MHz, CDC1₃): δ = 0.55-1.23 (CH₃ signals), 2.00 (s, OAc), 4.50-4.85 (m, 3-H), 5.11-5.25 (m, 7-H, 22-H, 23-H).- MS: m/z (%) = 456.3598 (50, M⁺, Calc for C₃₀H₄₈O₃ : 456.3603), 438 (28), 423 (4), 331 (16), 313 (14), 311 (8), 302 (100), 253 (32).

8,14-Epoxy-5a,8a-ergost-22-ene-38,7a-diol 3-acetate (42).

a) From 36: A solution of 36 (624.9 mg, 1.37 mmol) and pyridinium dichromate (747 mg, 1.99 mmol) in CH₂Cl₂ (60 ml) was stirred at 20°C for 7 h. Filtration through silica gel (elution with CH₂Cl₂ and ethyl acetate), solvent evaporation and LC (hexanes - ethyl acetate 7:1) gave 37 (292.3 mg, 28%) and 42 (339.1 mg, 41%).-M.p. 122-124°C (from methanol).- IR (CCl₄): 3575 (OH), 1730 cm⁻¹ (CO).- ¹H NMR (80 MHz, CDCl₃): & = 0.80-1.07 (CH₃ signals), 2.02 (s, OAc), 2.29 (m, OH), 3.56 (m, W1/2=5 Hz, 7-H), 4.55-5.00 (m, 3-H), 5.15-5.32 (m, 22-H, 23-H).- MS m/z (%) = 472 (22, M*), 457 (14), 379 (7), 356 (8), 347 (20), 329 (59), 311 (29), 269 (18), 251 (27), 236 (35), 107 (100).-(Found: C, 76.30; H, 10.18; CsoH4sO4 (427.7) requires C, 76.23; H, 10.24).

b) From 35: To a solution of 35 (20.0 mg, 0.044 mmol) in diethylether (1 ml) at -78°C a solution of MCPBA (13 mg, 0.064 mmol) in diethylether (0.5 ml) was added. The mixture was stirred at -78°C for 4 h and was then kept at -18°C for 44 h and at 4-6°C for 10 d. Usual work-up (diethylether) and LC (hexanes-ethyl acetate 5:1) gave 42 (9.7 mg, 47 %).

<u>38-Acetoxy-5g-ergosta-8(14),22-dien-7-one (37).</u>

M.p. 149-151°C (from methanol).- IR (CCl4): 1730, 1675 (CO), 1590 cm⁻¹ (C=C).- ¹H NMR (80 MHz, CDCl3): 8 = 0.80-1.00 (CH3 signals), 2.03 (OAc), 4.50-5.00 (3-H), 5.15-5.31 (22-H and 23-H).- MS: m/z (%) = 454 (M⁺, 72), 439 (20), 411 (7), 355 (18), 328 (100), 315 (18), 304 (14), 301 (11), 276 (15).- (Found: C, 79.19; H, 10.14; C30H46O3 (454.7) requires C, 79.25; H, 10.20).

8,14-Epoxy-5g,8g-ergost-22-ene-38,7g-diol_3-acetate 7-mesylate (43).

43 was prepared from 42 as described for 22 yield: 87%.- M.p. (dec.) 128°C (from

hexanes-CH₂Cl₂).- IR (CCl₄): 1730 (CO), 1360, 1170 cm⁻¹ (SO₂).- ¹H NMR (80 MHz, CDCl₃): $\delta = 0.80-1.06$ (CH₃ signals), 2.03 (s, OAc), 3.12 (s, SO₂CH₃), 4.61 (m, W_{1/2}=5 Hz, 7-H), 4.50-4.90 (m, 3-H), 5.17-5.31 (m, 22-H, 23-H).- MS: m/z (%) = 550 (2, M⁺), 454 (22), 439 (9), 436 (7), 425 (11), 411 (16), 383 (15), 337 (21), 329 (55), 328 (39), 311 (17), 107 (100).- (Found: C, 67.67; H, 9.04; C₃₁H₅₀O₆S (550.8) requires C, 67.60; H, 9.15).

8.14-Epoxy-5g.8g-ergosta-6.22-dien-38-yl acetate (44).

44 was prepared from 43 as described for 6 (44 h at 80° C): yield 72% .- M.p. 134-136°C (from methanol).- IR (CCl₄): 1730 cm⁻¹ (CO).- ¹H NMR (80 MHz, CDCl₃): $\delta = 0.80-1.07$ (CH₃ signals), 2.04 (s, OAc), 4.56-5.00 (m, 3-H), 5.13-5.37 (m, 6-H, 22-H, 23-H), 5.63 (A-part of an ABX-system, JAB =10 Hz, JAX=1.8 Hz, 7-H).- MS: m/z (%) = 454(1, M*), 436 (4), 394 (6), 376 (4), 361 (7), 329 (100), 311 (10), 269 (9), 251 (36), 235 (23), 175 (45), 107 (88).- (Found: C, 79.18; H, 10.25; C₃0H₄₆O₃ (454.7) requires C, 79.25; H, 10.20).

<u>38-Acetoxy-14-hydroxy-5g-ergosta-7.22-dien-6-one (38).</u>

a) From 33: 33 (prepared from 32 as described by Laubach et al. ⁹) was oxidized to give 38 as described for 7 : yield 48% - M.p. 194-195°C (from methanol).- IR (CHCl₃): 3600-3300 (OH), 1720 (CO), 1670 cm⁻¹ (unsat. CO).- ¹H NMR (80 MHz, CDCl₃): $\delta = 0.68$ (s, CH₃-18), 0.78-1.05 (CH₃ signals), 2.02 (s, OAc), 4.45-4.95 (m, 3-H), 5.13-5.28 (m, 22-H, 23-H), 5.89 (d, J=2.8 Hz, 7-H).- MS: m/z (%) = 470 (1, M⁺), 452 (2), 437 (5), 410 (100), 367 (8), 328 (12), 318 (11), 276 (49), 257 (50), 229 (39), 216 (65), 215 (41).- (Found: C, 76.43; H, 9.92; C₃0H46O4 (470.7) requires C,76.55; H, 9.85).

b) From 44: 44 was oxidized with sodium dichromate to give 38 as described for the formation of 7 from 6. The reaction was performed only in a 1 mg scale. TLC (hexanes-ethyl acetate 1:3) indicated the formation of a single product identical with 38 prepared from 33.

38.14-Dihydroxy-5g-ergosta-7.22-dien-6-one (39).

A mixture of 38 (290 mg, 0.62 mmol) in THF (15 ml) and 2.5 % K₂CO₃ in 1:1 methanol-water (20 ml) was stirred for 24 h at 25°C. The mixture was diluted with water, and methanol and THF were evaporated. Then usual work-up (diethylether) and LC (hexanes-ethyl acetate 1:2) gave 39 (198 mg, 75 %).- M.p. 219-222°C (from methanol).- IR (CHCl₃): 3600, 3550-3300 (OH), 1665 cm⁻¹ (unsat. CO).- ¹H NMR (80 MHz, CDCl₃): $\delta = 0.71$ (s, CH₃-18), 0.80-1.05 (CH₃ signals), 3.40-3.85 (m, 3-H), 5.18-5.32 (m, 22-H, 23-H), 5.91 (d, J=2.7 Hz, 7-H).- MS: m/z (%) = 430 (12), 429 (22), 428 (11, M⁺), 396 (10), 395 (11), 367 (6), 285 (15), 276 (48), 275 (33), 235 (98), 234 (100).- (Found: C, 78.33; H, 10.41; C2*H4*O3 (428.7) requires C, 78.46; H, 10.35).

14-Hydroxy-38-mesyloxy-5g-ergosta-7.22-dien-6-one (40).

39 was mesylated to furnish **40** as described for **30** : yield 78 - M.p. 195-196°C (from hexanes-CH₂Cl₂).- IR (CHCl₃): 3580-3300 (OH), 1665 (unsat. CO), 1350, 1170 cm⁻¹ (SO₂).- ¹H NMR (80 MHz, CDCl₃): δ = 0.70 (s, CH₃-18), 0.80-1.06 (CH₃ signals), 3.04 (s, SO₂CH₃), **4.45-4.90** (3-H), 5.15-5.32 (m, 22-H, 23-H), 5.92 (d, J=2.7 Hz, 7-H).- MS: m/z (%) = 506.3068 (2, M*, Calc for C₂₉H₄₆O₅S : 506.3066), 488 (11), 473 (14), 411 (17), 410 (23), 392 (32), 377 (38), 362 (51), 267 (47), 266 (56), 257 (28), 251 (34), 216 (44), 69 (100).

Reaction of 10 with butyllithium.

To a solution of 10 (200 mg, 0.46 mmol) in THF (5 ml) and HMPA (0.56 ml) at 0°C 1.6 M BuLi in hexane (2 ml, 3.2 mmol) was added, and the mixture was stirred at 20°C for 70 h. Usual work-up (diethylether) and MPLC (hexanes - ethyl acetate - methanol 10:10:0.75, then 10:10:1) furnished 5α -cholest-8(14)-ene-38,7 α -diol (45)

(65 mg, 35%, m.p. 154-156 °C (from methanol - water), lit.,¹² 157-158 °C, spectroscopic results in agreement with the published data ⁴²), and 46 (71 mg, 38%).

5a-Cholest-8-ene-38.7a-diol (46).

M.p. $133-135^{\circ}C$ (from methanol - water).- IR (CHCl₃): 3600, 3500-3300 cm⁻¹ (OH).-¹H NMR (80 MHz, CDCl₃): $\delta = 0.60$ (s, CH₃-18), 0.80-1.00 (CH₃ signals), 3.45-3.90 (3-H), 4.02 (m, W_{1/2}=6 Hz, 7-H).- MS: m/z (%) = 402 (M⁺, 0.5), 384 (100), 369 (22), 351 (14), 271 (20).- (Found: C, 80.36; H, 11.55; C₂₇H₄₆O₂ (402.6) requires C, 80.54; H, 11.52).

5a-Cholest-8-ene-3,7-dione (50).

A solution of 46 (40 mg, 0.1 mmol) and pyridinium dichromate (76 mg, 0.2 mmol) in CH_2Cl_2 (2 ml) was stirred at 20°C for 48 h. Filtration through silica gel (elution with CH_2Cl_2), solvent evaporation, and MPLC (hexanes - ethyl acetate 5:1) furnished 50 (32 mg, 82%).- M.p. 197-200°C (from methanol).- IR (CCl₄): 1720, 1670 cm⁻¹ (CO).- ¹H NMR (80 MHz, CDCl₃): δ = 0.61 (s, CH₃-18), 0.80-1.00 (CH₃ signals), 1.32 (s, CH₃-19).- MS: m/z (%) = 398.3176 (69, M*, Calc for C₂7H₄2O₂: 398.3185), 383 (14), 313 (4), 285 (44), 274 (16), 258 (26), 243 (39), 230 (62), 121 (100).

Acknowledgements- We wish to thank Prof.Dr.G.Snatzke for the CD spectra, Dr.W. Dietrich, Dr.D.Müller and their staffs (Ruhr-Universität Bochum, Lehrstuhl für Analytische Chemie) for the NMR and mass spectra, respectively. Financial support by the Deutsche Forschungsgemeinschaft and the Fonds der Chemischen Industrie is gratefully acknowledged.

References and Notes

- ¹ Review: D.Onken and D.Onken, Die Pharmazie 35, 193 (1980).
- ² c.f. P.Welzel, K.Hobert, A.Ponty, D.Neunert, and H.Klein,
- Tetrahedron, 41, 4509 (1985).
- ³ c.f. M.Anastasia, A.Fiecchi, A.Scala, and M.del Puppo, J.Chem.Soc., Perkin Trans I 1981, 2561; T.Matsumoto, J.Hasegawa, N.Shimizu, and T.Akihisa, Yakugaku Zasshi 34, 476 (1985)
- ⁴ c.f. T.Akihisa, P.Ghosh, S.Thakur, F.U.Rosenstein, and T.Matsumoto, J.Am.Oil Chem.Soc. 63, 653 (1986); Th.A.Salt and J.H.Adler, Lipids 21, 754 (1986); S.Xu, G.W.Patterson, and K.Schmid, Phytochemistry 25, 1883 (1986).
- ⁵ c.f. T.Rezanka, O.Vyhnalek, and M.Podojil, Folia Microbiol. (Prague) 31, 44 (1986), T.S.Milkova, S.S.Popov, N.L.Marekov, H.V.Dilov, and S.S.Yordonova, Compt.Rend.Acad.Bulg.Sci. 30, 1027 (1977).
- ⁶ For leading references, see I.L.Stoilov, M:Bladocha-Moreau, J.E.Thompson, and C.Djerassi, Tetrahedron 43, 2213 (1987); V.A.Stonik, Pure Appl.Chem. 58, 423 (1986).
- ⁷ U.Hedtmann and P.Welzel, Tetrahedron Lett. 26, 2773 (1985).
- ⁸ K.Wada, Agr.Biol.Chem. **39**, 1679 (1973).
- 9 G.D.Laubach, E.C.Schreiber, E.J.Agnello, and K.J.Brunings, J.Am.Chem.Soc. 78, 4743 (1956).
- 10 D.H.R.Barton, S.G.Davies, and W.B.Motherwell, Synthesis 1979, 265.
- 11 For an alternative approach, see D.H.R.Barton, P.G.Feakins, J.P.Poyser, and P.G.Sammes, J.Chem.Soc.(C) 1970, 1584.
- 12 L.F.Fieser and G.Ourisson, J.Am.Chem.Soc. 75, 4404 (1953).
- 13 L.F.Fieser and T.Goto, J.Am.Chem.Soc. 82, 1693 (1960).
- ¹⁴ c.f. I.Midgley and C.Djerassi, Tetrahedron Lett. 1972, 4673; B.N.Lutzky, J.A.Martin, and G.Schroepfer, Jr., J.Biol.Chem. 246, 6737 (1971).
- 15 U.Hedtmann, Dissertation, Universität Bochum 1987.
- 16 B.M.Trost, M.Lautens, and B.Peterson, Tetrahedron Lett. 24, 4525 (1983).
- ¹⁷ L.F.Fieser, K.Nakanishi, and Wei-Yuan Huang, J.Am.Chem.Soc.75, 4719 (1953).
- 18 I.Midgley and C.Djerassi, J.Chem.Soc., Perkin Trans. I 1972, 2771, and ibid. 1973, 155.

- ¹⁹ For a related case, see A.K.Sen Gupta, Chem. Ber. **100**, 694 (1967).
- ²⁰ Review: R.H.Shapiro, Org.React. 23, 405 (1976).
- ²¹ R.E.Ireland, D.C.Muchmore, and U.Hengartner, J.Am.Chem.Soc. 94, 5098 (1972).
- 22 Assuming a 100% yield for the acetylation step 5 6, which we did not perform.
- ²³ Prepared using the Canonica method.²⁴ For references, see "Rodd's Chemistry of Carbon Compounds", ed.S.Coffey, Academic Press, 2nd edition, 1970, vol.2, part D, p.85.
- ²⁴ L.Canonica, A.Fieci, M.Galli-Kienle, A.Scala, G.Galli, E.Grossi-Paoletti, and R.Paoletti, Steroids 11, 287 (1968).
- ²⁵ T.Milkova, unpublished results.
- ²⁶ Using the experimental results described by V.S.Jorapurand A.M.Shaligram, Indian J.Chem. **19b**, 940 (1980).
- ²⁷ c.f. A.Furlenmeier, A.Furst, A.Langemann, G.Waldvogel, P.Hocks, U.Kerb, and R.Wiechert, Helv.Chim. Acta 50, 2387 (1967), and ref.¹¹
- ²⁸ A.J.Pearson, Y.-S.Chen, G.R.Han, S.Y.Hsu, and T.Ray, J.Chem.Soc., Perkin Trans. I 1985, 267.
- ²⁹ For low yield oxidations, see J.K.Kinnear, M.-D.Martin, D.H.S.Horn, E.J.Middleton, J.S.Wilkie, M.N.Galbraith, and R.I.Willing, Austr.J.Chem. **29**, 1815 (1976).
- ³⁰ W.G.Salmond, M.A.Barta, and J.L.Havens, J.Org.Chem. 43, 2057 (1978).
- ³¹ Review: J.K.Crandall and M.Apparu, Org.React. 29, 345 (1983).
- 32 W.G.Dauben and D.M.Michno, J.Org.Chem. 42, 682 (1977).
- ³³ c.f. G.O.Schenk, W.Eisfeld, and O.A.Neumüller, Liebigs Ann. Chem. **1975**, 701.
- ³⁴ E.F.Parish, T.E.Spike, and G.J.Schoepfer, Jr., Chem. Phys.Lipids 18, 233 (1972), and references therein.
- 35 Prepared as described by D.Seebach and D.Enders, Chem.Ber. 108, 1293 (1975).
- ³⁶ J.Y.C.Chu, J.Chem.Soc., Chem.Commun. **1974**, 374, and references therein.
- ³⁷ F.De Simone, A.Dini, E.Finamore, L.Minale, C.Pizza, R.Riccio, and F.Zollo, J. Chem. Soc., Perkin Trans I 1981, 1855.
- ³⁸ c.f. M.Kobayashi, R.Tsuru, K.Todo, and H.Mitsuhashi, Tetrahedron 29, 1193 (1973).
- ³⁹ G.Sauci, P.Geistlich, R.Helbling, and H.Heusser, Helv. Chim. Acta 37, 250 (1954).
- 40 41 is possibly identical with the "9α-OH compound" described in ref.³⁹ Our structural assignment is based on the observation that 42 slowly rearranged into 41.⁴¹ No experiments to prove the structure unequivocally were performed.
- ⁴¹ c.f. the mechanistic discussion in ref.¹²
- ⁴² M.Tsuda, E.J.Parish, and G.J.Schroepfer, Jr., J. Org. Chem. 44, 1282 (1979).